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Synthesis of ²H- and ¹³C-labelled sunitinib and its primary metabolite

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Sunitinib (Sutent[®], Pfizer) was approved in 2006 for the treatment of gastrointestinal and renal cancer. Isotope-labelled derivatives have already been prepared for PET and ADME radiography. The preparation of ¹³C- and ²H-labelled internal standards of sunitinib (SU11248) and its primary metabolite (SU12662) for LC-MS analysis of human blood samples is presented.

Keywords: sunitinib; carbon-13; deuterium; SU11248; SU12662

Introduction

The discovery of SU11248, later named sunitinib, was published and patented by Sugen/Pharmacia in 2001 (Figure 1).^{1,2} In 2003 Pfizer acquired Sugen and by January 2006 sunitinib was approved by the US FDA for the treatment of gastrointestinal stromal tumours and advanced renal cell carcinoma.³ Marketed as Sutent[®], sunitinib is a multitargeted tyrosine kinase inhibitor with antiangiogenic and antitumour activities. It was selected from several oxindol derivatives as a selective inhibitor of the receptor tyrosine kinases VEGFR1-3, PDGFR α and β , KIT, FIt3, RET, and the CSF1 receptor.⁴ Sunitinib was found to be significantly more potent against VEGFR2 and PDGFR α than other candidate drugs and exhibited superior pharmacokinetic properties.⁵ Metabolism studies revealed that sunitinib is mainly metabolized by *N*-desethylation through CYP3A4 to the still active metabolite SU12662 as shown in Figure 1.⁶

Identification and quantification of drugs and metabolites by LC/MS relies very much on stable isotope-labeled analogues.^{7,8} Analytical procedures to determine both compounds, SU11248 and SU12662, in biological matrices, e.g. monkey blood samples, have been published.⁹ Unfortunately, the internal standard and its synthesis used in this study was not completely specified by the authors. One might conclude from the details provided (m/z = 409, most probably ESI⁺), that a sunitinib- d_{10} analogue was used. In addition, ¹⁸F-sunitinib has been prepared for PET purposes and a ¹⁴C-analogue was synthesized for ADME studies prior to FDA approval.^{10,11} The aim of our work was to prepare comparable ¹³C/²H-SU11248/SU12662 internal standards for LC-MS determination of sunitinib and its primary metabolite in human blood samples.

Results and discussion

N-Alkylation, e.g. *N*-*d*₅-ethylation, has been used to stable label a variety of drugs to obtain internal standards for biological quantification.^{12–14} Between the sunitinib-*d*₅, presented herein, and sunitinib itself, neither cross-contribution to analyte ions⁷ nor any difference in chromatographic behaviour,¹⁵ e.g. lower



N-desethyl-sunitinib (SU12662)

Figure 1. Sunitinib is mainly metabolized by CYP3A4 through desalkylation of the basic side chain to provide *N*-desethyl-sunitinib, which is still active.

retention times, was observed (see Supporting Information, S9, S12). Their physicochemical properties appeared therefore

Supporting information may be found in the online version of this article.

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very similar and the internal standard was considered suitable for the determination of sunitinib. Next to the presented synthesis of a sunitinib- d_5 internal standard, an analogously d_5 -deuterized metabolite, requiring mono-alkylation of ethylenediamine, posed a major chemical challenge, and was therefore not addressed. Nevertheless, also SU12662 was successfully analysed using the sunitinib- d_5 internal standard. Structural elucidation of the sunitinib scaffold was based particularly on NMR experiments exploiting the unique ¹H, ¹³C, and ¹⁹F coupling patterns observed with the prepared ¹³C-isotopomers.

The convergent synthesis of sunitinib and related indolin-2one derived structures started from 5-fluoroindolin-2-one (1), which was readily obtained by a Wolff-Kishner-type reduction of the corresponding 5-fluoroisatin (Scheme 1). The pyrrolecontaining building block was prepared from commercially available ethyl 2,4-dimethyl-1H-pyrrole-3-carboxylate (Ace Synthesis, Woburn, USA). A Vilsmeier reaction, introducing a ¹²C- or ¹³C-formyl function into the pyrrole, was carried out affording compounds 2a and 2b. Subsequent alkaline hydrolysis of the ethyl ester moiety produced 3a and 3b, which where joined with 1 in a pyrrolidine-catalyzed aldol reaction to obtain key intermediates 4a and 4b. Reacting ethyl esters 2 with 1 prior to alkaline hydrolysis is not advisable, as the poor solubility of 3-substituted indolin-2-ones, e.g. 4, impairs following reaction steps. Compounds 4a and 4b were obtained exclusively in Z-configuration, a fact that has already been investigated in detail.¹⁶ The mono- or dialkylaminoethylcarboxamide side chains of sunitinib (¹³C: 5) and its primary metabolite (¹²C: 6a, ¹³C: **6b**) where introduced in a final step through an EDC/HOBT coupling procedure applying careful temperature control, i.e. lowered reaction temperature in case of *N*-ethylethylenediamine to receive regioselectivity in favour of the primary amino function. No side chain precursor was commercially available for the synthesis of the sunitinib- d_5 internal standard. Therefore, N-ethylethylenediamine was reacted in a Gabriel-like synthesis to the phthalimide 7, which was subsequently alkylated with [²H₅]ethyl iodide (Scheme 2). Final hydrazinolysis provided the deuterized side chain precursor N,N-[²H₅]diethylethylenediamine in situ, which was reacted with **4a** to afford sunitinib- d_5 (8). Purification of 5, 6a, 6b, and 8 was achieved by preparative column chromatography. The corresponding salts, i.e. 5 and 6b, were obtained by lyophilization from dilute hydrochloric acid.

Experimental section

General methods and materials

Melting points were determined on a Boëtius melting point apparatus (VEB Wägetechnik Rapido PHMK) and are uncorrected. Thin-layer chromatography was performed on aluminium sheets coated with silica gel 60 F₂₅₄, preparative column chromatography using silica gel 60, 70-230 mesh (Merck, Darmstadt, Germany) or a C₁₈ column (Knauer Eurospher 100, $10\,\mu m$, $250\,mm \times 20\,mm$). LC-MS analysis was performed on an API 2000 mass spectrometer (ESI, Applied Biosystems, Darmstadt, Germany), coupled with an HPLC system (Agilent 1100) using a C_{18} column (Phenomenex Luna, $3\,\mu\text{m},~50\,{\times}\,2\,\text{mm},$ Aschaffenburg, Germany) and a gradient of MeOH/H₂O. Additional MS² experiments were carried out to acquire the corresponding fragmentation patterns. HPLC purity assessment was carried out using a C18 column (Ziemer Hypersil-ODS, $3 \,\mu$ m, $125 \times 4.6 \,m$ m, Langerwehe, Germany) attached to a UV detector (Jasco UV-2075Plus, 220 nm) under isocratic conditions



Scheme 1. Synthesis of ^{12/13}C-labelled precursors, sunitinib and its primary metabolite. (a) N₂H₄, Et₃N, *n*-BuOH, 100°C, 16 h; (b) 1. $[1^{12}C/^{13}C]DMF$, POCl₃, CH₂Cl₂, 4°C, 15 min to reflux, 1 h; 2. HCl, H₂O, RT, 30 min; (c) KOH, H₂O, reflux, 5 h; (d) EtOH, pyrrolidine (cat.), reflux, 3 h; (e) H₂N(CH₂)₂NEt₂, HOBT, EDC, Et₃N, DMF, RT, 48 h; (f, g) H₂N(CH₂)₂NHEt, HOBT, EDC, Et₃N, DMF, -60°C, 6 h to RT, 18 h.

(MeCN/H₂O, 65/35 v/v). ¹³C NMR (125 MHz) and ¹H NMR spectra (500 MHz) were recorded on a Avance DRX 500 spectrometer (Bruker BioSpin, Rheinstetten, Germany) and chemical shifts δ are given in ppm referring to the signal centre using the solvent peaks for reference (DMSO-*d*₆: 2.49/39.7). To characterize the spin multiplicity the following abbreviations are used: s singlet, bs broad singlet, d doublet, dd doublet of doublets, ddd doublet of doublet of doublet, t triplet, q quartet, dq doublet of quartets, m multiplet. Apparent spin multiplicity is denoted by a preceding ,app'. ¹³C NMR signals were assigned on the basis of ¹³C/¹⁴F, ¹³C/¹⁹F, and ¹³C/¹³C coupling patterns.

5-Fluoro-1,3-dihydro-2H-indol-2-one (1)

5-Fluoroisatin (30.0 mmol, 4.95 g) and hydrazine (61.8 mmol, 3.0 mL hydrazine hydrate, 100%) were suspended in *n*-butanol (50 mL) and stirred at room temperature for 30 min.¹⁷ After heating to 80° C for 3 h, triethylamine (5 mL) was added and the suspension was stirred further 12 h at 100° C. Upon cooling to room temperature and solvent removal *in vacuo* the crude product was dissolved in ethyl acetate (100 mL) and washed



Scheme 2. Synthesis of ²H-labelled sunitinib. (a) $H_2N(CH_2)_2NHEt$, $100^{\circ}C$, 3 h; (b) $[^{2}H_{3}]EtI$, K_2CO_3 , MeCN, RT, 16 h; (c) N_2H_4 , EtOH, reflux, 1 h; (d) HOBT, EDC, Et_3N, DMF, RT, 18 h.

with 10% potassium hydrogen sulphate solution. The aqueous layer was extracted with ethyl acetate $(1 \times 100 \text{ mL})$, the combined organic phases were washed with brine $(1 \times 50 \text{ mL})$ and evaporated in vacuo. The residue was dissolved in hot ethyl acetate (50 mL) and petroleum ether was added until the solution turned slightly cloudy. After filtration and cooling to room temperature, 1 was separated by suction filtration to afford light brown crystals (3.12 g, 69%), mp 128°C, lit.¹⁸ 121–134°C. ¹H NMR (DMSO-*d*₆) δ: 3.47 (s, 2H, 3-H), 6.76 (dd, 1H, ${}^{3}J$ (H,H) = 8.5 Hz, ${}^{4}J$ (H,F) = 4.7 Hz, 7-H), 6.97 (ddd, 1H, ${}^{3}J$ (H,F) = 9.1 Hz, ${}^{3}J(H,H) = 8.3 Hz$, ${}^{4}J(H,H) = 2.9 Hz$, 6-H), 7.07 (dd, 1H, ${}^{3}J$ (H,F) = 8.6 Hz, ${}^{4}J$ (H,H) = 2.8 Hz, 4-H), 10.32 (s, 1H, NH); ${}^{13}C$ NMR (DMSO- d_6) δ : 36.32 (C-3), 109.70 (d, 1C, ³J (C,F) = 8.4 Hz, C-7), 112.32 (d, 1C, ${}^{2}J$ (C,F) = 24.5 Hz, C-6), 113.67 (d, 1C, ${}^{2}J$ (C,F) = 23.1 Hz, C-4), 127.81 (d, 1C, ³J (C,F) = 8.9 Hz, C-3a), 140.05 (C-7a), 157.87 (d, 1C, ¹J (C,F) = 234 Hz, C-5), 176.32 (C-2). HPLC purity: 99.9%.

Ethyl 5-formyl-2,4-dimethyl-1H-pyrrole-3-carboxylate (2a)

A solution of *N*,*N*-dimethyl-formamide (11.0 mmol, 0.80 g) and phosphorus(V) oxychloride (11.0 mmol, 1.69 g) in dichloromethane (10 mL) was cooled to 4°C and solid ethyl 2,4-dimethyl-1*H*-pyrrole-3-carboxylate (10.0 mmol, 1.67 g) was added slowly (maximum temperature 10°C). The reaction mixture was stirred 15 min and subsequently heated to reflux for 1 h. Upon cooling to 10°C water (5 mL), followed by hydrochloric acid (10 M, 5 mL) was added with vigorous stirring. The layers were allowed to separate and the organic phase was extracted with hydrochloric acid (10 M, 2 × 10 mL). The combined aqueous extracts were washed with dichloromethane (1 × 20 mL), and sodium hydroxide (10 M, 25 mL) was added to afford **2a** as a yellow precipitate (1.89 g, 97%), mp 167°C, lit.¹⁹ 163–164°C. ¹H NMR (DMSO- d_6) δ : 1.26 (t, 3H, ³*J* (H,H) = 7.1 Hz, CH₂CH₃), 2.41 (s, 3H, 2-CH₃), 2.45 (s, 3H, 4-CH₃), 4.18 (q, 2H, ³*J* (H,H) = 7.2 Hz, CH₂), 9.60 (s, 1H, CHO), 12.13 (s, 1H, NH); ¹³C NMR (DMSO- d_6) δ : 10.48 (4-CH₃), 13.66 (2-CH₃), 14.38 (CH₂CH₃), 59.18 (CH₂), 112.81 (C-3), 128.41 (C-5), 133.80 (C-4), 142.73 (C-2), 164.45 (CO₂CH₂CH₃), 178.01 (CHO). HPLC purity: 98.9%.

Ethyl 5-[¹³C]formyl-2,4-dimethyl-1*H*-pyrrole-3-carboxylate (2b)

N,*N*-dimethyl-[¹³C]formamide (11.0 mmol, 0.82 g) was reacted as described above to afford **2b** as a yellow precipitate (1.78 g, 91%), mp 158°C. ¹H NMR (DMSO-*d*₆) δ : 1.27 (t, 3H, ³*J* (H,H) = 7.1 Hz, CH₂CH₃), 2.41 (s, 3H, 2-CH₃), 2.45 (s, 3H, 4-CH₃), 4.18 (q, 2H, ³*J* (H,H) = 7.1 Hz, CH₂), 9.60 (d, 1H, ¹*J* (H,C) = 174 Hz, CHO), 12.12 (s, 1H, NH); ¹³C NMR (DMSO-*d*₆) δ : 10.48 (4-CH₃), 13.66 (2-CH₃), 14.38 (CH₂CH₃), 59.19 (CH₂), 112.81 (d, 1C, ³*J* (C,C) = 3.7 Hz, C-3), 128.39 (d, 1C, ¹*J* (C,C) = 65.7 Hz, C-5), 133.81 (d, 1C, ²*J* (C,C) = 5.2 Hz, C-4), 142.72 (C-2), 164.45 (CO₂CH₂CH₃), 178.02 (CHO). HPLC purity: 100.0%.

5-Formyl-2,4-dimethyl-1H-pyrrole-3-carboxylic acid (3a)

A suspension of ethyl 5-formyl-2,4-dimethyl-1*H*-pyrrole-3-carboxylate (**2a**; 8.0 mmol, 1.56 g) in a solution of potassium hydroxide (90%, 16.0 mmol, 1.0 g) in water (18 mL) was heated to reflux for 5 h. After cooling to room temperature the clear solution was diluted with water (30 mL) and washed with dichloromethane (1 × 40 mL). Subsequently, the pH was adjusted to 4 using concentrated hydrochloric acid to recover **3a** as a yellow precipitate (1.19 g, 89%), mp 286°C (decomposition). ¹H NMR (DMSO-*d*₆) δ : 2.41 (s, 3H, 2-CH₃), 2.44 (s, 3H, 4-CH₃), 9.59 (s, 1H, CHO), 12.04 (s, 2H, CO₂H, NH); ¹³C NMR (DMSO-*d*₆) δ : 10.53 (4-CH₃), 13.72 (2-CH₃), 113.51 (C-3), 128.37 (C-5), 134.21 (C-4), 142.89 (C-2), 166.12 (CO₂H), 177.89 (CHO). HPLC purity: 99.4%.

5-[¹³C]Formyl-2,4-dimethyl-1*H*-pyrrole-3-carboxylic acid (3b)

A suspension of ethyl 5-[¹³C]formyl-2,4-dimethyl-1*H*-pyrrole-3carboxylate (**2b**; 8.0 mmol, 1.57 g) was treated as described above to recover **3b** as a yellow precipitate (1.11 g, 83%), mp 285°C (decomposition). ¹H NMR (DMSO- d_6) δ : 2.41 (s, 3H, 2–CH₃), 2.44 (s, 3H, 4–CH₃), 9.59 (d, 1H, ¹J (H,C) = 174 Hz, CHO), 12.04 (s, 2H, CO₂H, NH); ¹³C NMR (DMSO- d_6) δ : 10.52 (4-CH₃), 13.70 (2-CH₃), 113.51 (d, 1C, ³J (C,C) = 3.5 Hz, C-3), 128.34 (d, 1C, ¹J (C,C) = 65.9 Hz, C-5), 134.19 (d, 1C, ²J (C,C) = 5.2 Hz, C-4), 142.87 (d, 1C, ³J (C,C) = 2.5 Hz, C-2), 166.11 (CO₂H), 177.89 (CHO). HPLC purity: 99.9%.

5-((5-Fluoro-2-oxo-1,2-dihydro-3*H*-indol-3-ylidene)methyl)-2,4-dimethyl-1*H*-pyrrole-3- carboxylic acid (4a)

A solution of 5-fluoro-1,3-dihydro-2*H*-indol-2-one (**1**; 8.8 mmol, 1.33 g), 5-formyl-2,4-dimethyl-1*H*-pyrrole-3-carboxylic acid (**3a**; 8.8 mmol, 1.47 g), and pyrrolidine (18.0 mmol, 1.5 mL) in ethanol (120 mL) was heated to reflux for 3 h. Upon cooling to room temperature hydrochloric acid (2*M*, 15 mL) was added to the suspension, a crude precipitate was recovered by suction filtration and washed with ethanol (20 mL) followed by petroleum ether (20 mL) to afford **4a** as a yellow powder

(2.59 g, 98%), mp 320°C (decomposition). ¹H NMR (DMSO- d_6) δ : 2.49 (s, 3H, 2-CH₃), 2.52 (s, 3H, 4-CH₃), 6.83 (dd, 1H, ³J (H,H) = 8.5 Hz, ⁴J (H,F) = 4.8 Hz, 7'-H), 6.92 (ddd, 1H, ³J (H,F) = 9.5 Hz, ³J (H,H) = 8.5 Hz, ⁴J (H,H) = 2.5 Hz, 6'-H), 7.72 (s, 1H, 3'-CH-5), 7.74 (dd, 1H, ³J (H,F) = 9.5 Hz, ⁴J (H,H) = 2.5 Hz, 4'-H), 10.88 (s, 1H, NH_{ind}), 12.07 (s, 1H, CO₂H), 13.84 (s, 1H, NH_{pyr}); ¹³C NMR (DMSO- d_6) δ : 11.59 (4-CH₃), 14.62 (2-CH₃), 106.32 (d, 1C, ²J (C,F) = 25.5 Hz, C-4'), 110.24 (d, 1C, ³J (C,F) = 8.4 Hz, C-7'), 112.85 (d, 1C, ²J (C,F) = 24.0 Hz, C-6'), 114.53 (C-3), 115.83 (d, 1C, ⁴J (C,F) = 3.0 Hz, C-3'), 124.90 (3'-CH-5), 126.21 (C-5), 127.15 (d, 1C, ³J (C,F) = 9.4 Hz, C-3a'), 133.58 (C-4), 134.89 (C-7a'), 141.02 (C-2), 158.42 (d, 1C, ¹J (C,F) = 233 Hz, C-5'), 166.06 (CO₂H), 169.76 (C-2'). HPLC purity: 97.3%.

5-((5-Fluoro-2-oxo-1,2-dihydro-3*H*-indol-3-ylidene)-[¹³C]methyl)-2,4-dimethyl-1*H*-pyrrole-3-carboxylic acid (4b)

A solution of 5-fluoro-1,3-dihydro-2H-indol-2-one (1; 3.0 mmol, 0.45 g), 5-[¹³C]formyl-2,4-dimethyl-1*H*-pyrrole-3-carboxylic acid (3b; 3.0 mmol, 0.59 g), and pyrrolidine (6.0 mmol, 0.5 mL) in ethanol (20 mL) was reacted as described before to obtain 4b as a yellow powder (0.86 g, 95%), mp 316°C (decomposition). ¹H NMR (DMSO-d₆) δ: 2.49 (s, 3H, 2-CH₃), 2.52 (s, 3H, 4-CH₃), 6.83 $(dd, 1H, {}^{3}J(H,H) = 8.2 Hz, {}^{4}J(H,F) = 4.4 Hz, 7'-H), 6.91 (ddd, 1H, {}^{3}J$ (H,F) = 9.0 Hz, ³J(H,H) = 9.0 Hz, ⁴J(H,H) = 2.3 Hz, 6'-H), 7.70 (d, 1H, ^{1}J (H,C) = 153 Hz, 3'-CH-5), 7.73 (dd, 1H, ^{3}J (H,F) = 9.2 Hz, ^{4}J (H,H) = 2.2 Hz, 4'-H), 10.89 (s, 1H, NH_{ind}), 13.82 (s, 1H, NH_{pvr}); ¹³C NMR (DMSO-*d*₆) δ: 11.59 (4-CH₃), 14.62 (2-CH₃), 106.26 (d, 1C, ²J (C,F) = 25.3 Hz, C-4'), 110.22 (d, 1C, ³J (C,F) = 8.4 Hz, C-7'), 112.78 (d, 1C, ${}^{2}J$ (C,F) = 24.0 Hz, C-6'), 114.98 (C-3), 115.63 (dd, 1C, ${}^{1}J$ $(C,C) = 71.7 \text{ Hz}, {}^{4}J (C,F) = 2.5 \text{ Hz}, C-3'), 124.90 (3'-CH-5), 126.24 (d,$ 1C, ¹J (C,C) = 68.9 Hz, C-5), 127.18 (d, 1C, ³J (C,F) = 8.7 Hz, C-3a'), 133.60 (d, 1C, ${}^{2}J$ (C,C) = 4.5 Hz, C-4), 134.87 (d, 1C, ${}^{4}J$ (C,F) = 4.5 Hz, C-7a'), 140.97 (C-2), 158.41 (d, 1C, ¹J (C,F) = 233 Hz, C-5'), 166.25 (CO_2H) , 169.76 (d, 1C, ²J (C,C) = 2.5 Hz, C-2'). HPLC purity: 97.9%.

N-(2-(Diethylamino)ethyl)-5-((5-fluoro-2-oxo-1,2-dihydro-3*H*indol- 3 -ylidene)-[¹³C]methyl)-2,4-dimethyl-1*H*-pyrrole-3-carboxamide hydrochloride (5, ¹³C-SU11248)

A solution of 5-((5-fluoro-2-oxo-1,2-dihydro-3H-indol-3-ylidene)- $[^{13}C]$ methyl) - 2,4 - dimethyl - 1*H* - pyrrole - 3 - carboxylic acid (4; 1.0 mmol, 0.30 g), N,N-diethylethylenediamine (1.2 mmol, 0.14 g), 1-hydroxybenzotriazole (1.5 mmol, 0.20 g), 1-(3-dimethylaminopropyl)-3-ethylcarbodiimide hydrochloride (2.0 mmol, 0.38 g), and triethylamine (2.0 mmol, 0.20 g) in anhydrous N,Ndimethylformamide (10 mL) was stirred at room temperature for 48 h. The reaction mixture was subsequently evaporated in vacuo and the crude residue subjected to preparative HPLC on C₁₈-silica using a gradient of methanol–water containing 1.5% triethylamine. The product fractions were combined, evaporated in vacuo and lyophilized from hydrochloric acid (0.1 M, 25 mL) to obtain 5 as an orange powder (0.40 g, 91%), mp 274°C (decomposition). ¹H NMR (DMSO- d_6) δ : 1.25 (t, 6H, ³J (H,H) = 7.3 Hz, CH₂CH₃), 2.45 (s, 3H, 2-CH₃), 2.47 (s, 3H, 4-CH₃), 3.15 (dq, 4H, ${}^{3}J$ (H,H) = 4.8 Hz, ${}^{3}J$ (H,H) = 7.3 Hz, CH₂CH₃), 3.20 $(app q, 2H, {}^{3}J (H,H) = 6.1 Hz, CONHCH_{2}CH_{2}), 3.61 (app q, 2H, {}^{3}J)$ $(H,H) = 6.3 \text{ Hz}, \text{ CONHCH}_2\text{CH}_2), 6.84 \text{ (dd, } \overline{1H}, {}^3J \text{ (H,H)} = 8.5 \text{ Hz}, {}^4J$ $(H,F) = 4.7 Hz, 7'-H), 6.91 (ddd, 1H, {}^{3}J (H,F) = 9.5 Hz, {}^{3}J$ ¹J (H,H) = 8.5 Hz, ⁴J (H,H) = 2.6 Hz, 6'-H), 7.70 (d, 1H, $(H,C) = 153 Hz, 3'-CH-5), 7.74 (dd, 1H, {}^{3}J (H,F) = 9.5 Hz, {}^{4}J$ $(H,H) = 2.5 Hz, 4'-H), 8.02 (t, 1H, {}^{3}J (H,H) = 5.7 Hz, CONHCH₂CH₂),$ 8.03 (bs, 1H, N⁺H), 10.92 (s, 1H, NH_{ind}), 13.72 (s, 1H, NH_{pvr});

¹³C NMR (DMSO- d_6) δ : 8.61 (CH₂CH₃), 10.92 (4-CH₃), 13.74 (2-CH₃), 34.12 (CONHCH₂CH₂), 47.03 (CH₂CH₃), 50.37 (CONHCH₂CH₂), 106.15 (d, 1C, ²J (C,F) = 25.3 Hz, C-4'), 110.23 (d, 1C, ³J (C,F) = 8.4 Hz, C-7'), 112.62 (d, 1C, ²J (C,F) = 24.3 Hz, C-6'), 115.11 (d, 1C, ¹J (C,C) = 71.3 Hz, ⁴J (C,F) = 2.5 Hz, C-3'), 119.84 (d, 1C, ³J (C,C) = 3.0 Hz, C-3), 124.98 (3'-CH-5), 126.04 (d, 1C, ¹J (C,C) = 68.7 Hz, C-5), 127.26 (d, 1C, ³J (C,F) = 8.7 Hz, C-3a'), 130.58 (d, 1C, ²J (C,C) = 4.7 Hz, C-4), 134.78 (d, 1C, ⁴J (C,F) = 4.5 Hz, C-7a'), 137.16 (C-2), 158.40 (d, 1C, ¹J (C,F) = 233 Hz, C-5'), 165.37 (CONHCH₂CH₂), 169.72 (d, 1C, ²J (C,C) = 2.7 Hz, C-2'). LC-MS: purity 99.7%. MS²: ESI⁺ 400.1 ([C₂₁³CH₂₇FN₄O₂+H]⁺, 40%), 327.1 ([C₂₁³CH₂₇FN₄O₂-C₄H₁₀N]⁺, 52%), 283.9 ([C₂₁³CH₂₇FN₄O₂-C₆H₁₅N₂]⁺, 100%); ESI⁻ 398.1 ([C₂₁³CH₂₇FN₄O₂-H]⁻, 100%), 256.1 ([C₂₁¹³CH₂₇FN₄O₂-C₇H₁₅N₂O]⁻, 34%).

N-(2-(Ethylamino)ethyl)-5-((5-fluoro-2-oxo-1,2-dihydro-3*H*indol-3-ylidene)methyl)-2,4-dimethyl-1*H*-pyrrole-3-carboxamide (6a, SU12662)

A suspension of 5-((5-fluoro-2-oxo-1,2-dihydro-3H-indol-3-ylidene)methyl)-2,4-dimethyl-1H-pyrrole-3-carboxylic acid (4a; 2.0 mmol, 0.60 g), 1-hydroxybenzotriazole (3.0 mmol, 0.40 g), 1-(3-dimethylaminopropyl)-3-ethylcarbodiimide hydrochloride (4.0 mmol, 0.76 g), and triethylamine (6.0 mmol, 0.60 g) in N,N-dimethylformamide (20 mL) was stirred at room temperature for 2 h. Upon cooling to -60° C *N*-ethylethylenediamine (2.4 mmol, 0.22 g) in N,N-dimethylformamide (1 mL) was added, the solution was stirred 6 h at -60° C and warmed to room temperature within 18 h. Solvent removal provided a crude residue that was subjected to column chromatography on silica using a mixture of ethyl acetate, methanol, and triethylamine (8:2:1). Product fractions were combined, evaporated, dissolved in hydrochloric acid (1.0 M, 10 mL), and subsequently filtered. 6a was precipitated as a yellow powder (0.55 g, 74%) after addition of sodium hydroxide solution (2.0 M, 5 mL), mp 280°C (decomposition). ¹H NMR (DMSO- d_6) δ : 1.11 (t, 3H, ³J (H,H) = 7.1 Hz, CH₂CH₃), 2.43 (s, 3H, 2-CH₃), 2.45 (s, 3H, 4-CH₃), 2.76 (app g, 2H, ${}^{3}J$ (H,H) = 7.2 Hz, CH₂CH₃), 2.85 (app t, 2H, ${}^{3}J$ (H,H) = 6.5 Hz, CONHCH₂CH₂), 3.40 (bs, 1H, NH), 3.41 (app q, 2H, ${}^{3}J$ (H,H) = 6.1 Hz, CONHCH₂CH₂), 6.84 (dd, 1H, ${}^{3}J$ (H,H) = 8.4 Hz, ^{4}J (H,F) = 4.8 Hz, 7'-H), 6.91 (ddd, 1H, ^{3}J (H,F) = 9.5 Hz, ^{3}J (H,H) = 8.5 Hz, ⁴J (H,H) = 2.5 Hz, 6'-H), 7.70 (s, 1H, 3'-CH-5), 7.71 (t, 1H, ${}^{3}J$ (H,H) = 5.7 Hz, CONHCH₂CH₂), 7.74 (dd, 1H, ${}^{3}J$ (H,F) = 9.5 Hz, ${}^{4}J$ (H,H) = 2.5 Hz, 4'-H, 10.90 (s, 1H, NH_{ind}), 13.69 (s, 1H, NH_{pyr}); ¹³C NMR (DMSO-d₆) δ: 10.78 (4-CH₃), 13.26 (CH2CH3), 13.58 (2-CH3), 37.40 (CONHCH2CH2), 42.74 (CH2CH3), 47.53 (CONHCH₂<u>C</u>H₂), 106.07 (d, 1C, ²J (C,F) = 26.5 Hz, C-4'), 110.18 (d, 1C, ${}^{3}J$ (C,F) = 8.7 Hz, C-7'), 112.53 (d, 1C, ${}^{2}J$ (C,F) = 23.8 Hz, C-6'), 114.86 (d, 1C, ⁴J (C,F)=2.7 Hz, C-3'), 120.54 (C-3), 124.99 (3'-CH-5), 125.95 (C-5), 127.28 (d, 1C, ³J (C,F) = 9.4 Hz, C-3a'), 130.50 (C-4), 134.69 (C-7a'), 136.88 (C-2), 158.38 $(d, 1C, {}^{1}J, (C,F) = 233 \text{ Hz}, C-5')$, 165.12 (CONHCH₂CH₂), 169.70 (C-2'). LC-MS: purity 99.2%. MS²: ESI⁺ 371.1 ([C₂₀H₂₃FN₄O₂+H]⁺, 17%), 325.9 ([C₂₀H₂₃FN₄O₂-C₂H₆N]⁺, 10%), 282.9 ([C₂₀H₂₃FN₄O₂-C₄H₁₁N₂]⁺, 100%); ESI⁻ 368.9 ([C₂₀H₂₃FN₄O₂-H]⁻, 100%), 254.9 ([C₂₀H₂₃FN₄O₂-C₅H₁₁N₂O]⁻, 34%).

N-(2-(Ethylamino)ethyl)-5-((5-fluoro-2-oxo-1,2-dihydro-3*H*indol-3-ylidene)-[¹³C]methyl)-2,4-dimethyl-1*H*-pyrrole-3-carboxamide hydrochloride (6b, ¹³C-SU12662)

A suspension of 5-((5-fluoro-2-oxo-1,2-dihydro-3*H*-indol-3-ylidene)- $[^{13}C]$ methyl) - 2,4 - dimethyl - 1*H* - pyrrole - 3 - carboxylic acid (**4b**; 1.0 mmol, 0.30 g), 1-hydroxybenzotriazole (1.5 mmol, 0.20 g),

hydrochloride 1-(3-dimethylaminopropyl)-3-ethylcarbodiimide (2.0 mmol, 0.38 g), and triethylamine (3.0 mmol, 0.30 g) in N,Ndimethylformamide (20 mL) was stirred at room temperature for 2 h. Upon cooling to -60° C *N*-ethylethylenediamine (1.2 mmol, 0.11 g) in N,N-dimethylformamide (1 mL) was added, the solution was stirred 6 h at -60° C and warmed to room temperature within 18 h. Solvent removal provided a crude residue that was subjected to preparative HPLC on C18-silica using a gradient of methanolwater containing 1.5% triethylamine. Product fractions were combined, evaporated in vacuo and dissolved in a mixture of hydrochloric acid (0.1 M, 100 mL) and methanol (400 mL). Evaporation in vacuo afforded **6b** as a vellow powder (0.33 g, 81%), mp 306°C (decomposition). ¹H NMR (DMSO- d_6) δ : 1.21 (t, 3H, ³J (H,H) = 7.3 Hz, CH₂CH₃), 2.46 (s, 3H, 2-CH₃), 2.48 (s, 3H, 4-CH₃), 2.97 (app q, 2H, ${}^{3}J$ (H,H) = 7.1 Hz, CH₂CH₃), 3.05 (app t, 2H, ${}^{3}J$ $(H,H) = 6.2 \text{ Hz}, \text{ CONHCH}_2\text{CH}_2$, 3.55 (app q, 2H, ³J (H,H) = 6.1 Hz, CONHCH₂CH₂), 6.84 (dd, 1H, ${}^{3}J$ (H,H) = 8.5 Hz, ${}^{4}J$ (H,F) = 4.8 Hz, 7'-H), 6.92 (ddd, 1H, ³J (H,F) = 9.6 Hz, ³J (H,H) = 8.4 Hz, ⁴J $(H,H) = 2.6 Hz, 6'-H), 7.71(d, 1H, {}^{1}J(H,C) = 152 Hz, 3'-CH-5), 7.75$ (dd, 1H, ³J (H,F) = 9.5 Hz, ⁴J (H,H) = 2.5 Hz, 4'-H), 7.88 (t, 1H, ³J (H,H) = 5.7 Hz, CONHCH₂CH₂), 8.98 (bs, 2H, N⁺H₂), 10.90 (s, 1H, NH_{ind}), 13.72 (s, 1H, NH_{pyr}); ¹³C NMR (DMSO-*d*₆) δ: 10.89 (4-CH₃), 11.05 (CH₂<u>C</u>H₃), 13.75 (2-CH₃), 35.67 (CONH<u>C</u>H₂CH₂), 42.13 (CH_2CH_3) , 46.25 $(CONHCH_2CH_2)$, 106.10 $(d, 1C, {}^{2}J, (C,F) = 25.5 Hz$, \overline{C} -4'), 110.20 (d, 1C, ³J (C,F)=8.7 Hz, C-7'), 112.58 (d, 1C, ²J $(C,F) = 24.0 \text{ Hz}, C-6'), 115.00 (dd, 1C, {}^{1}J (C,C) = 71.3 \text{ Hz},$ ⁴J (C,F) = 2.7 Hz, C-3'), 119.92 (d, 1C, ³J (C,C) = 3.0 Hz, C-3), 124.96 (3'-CH-5), 126.02 (d, 1C, ¹J (C,C) = 68.9 Hz, C-5), 127.23 (d, 1C, ³J (C,F) = 9.4 Hz, C-3a'), 130.63 (d, 1C, ²J (C,C) = 4.7 Hz, C-4), 134.72 (d, 1C, ${}^{4}J$ (C,F) = 4.5 Hz, C-7a'), 137.22 (C-2), 158.37 (d, 1C, ${}^{1}J$ (C,F) = 233 Hz, C-5'), 165.43 (CONHCH₂CH₂), 169.70 (d, 1C, 2 J (C,C) = 2.7 Hz, C-2'). LC-MS: purity 99.3%. MS²: ESI⁺ 372.1 $([C_{19}^{13}CH_{23}FN_4O_2+H]^+, 16\%), 327.1 ([C_{19}^{13}CH_{23}FN_4O_2-C_2H_6N]^+, 13\%),$ 369.9 283.9 $([C_{19}^{13}CH_{23}FN_4O_2-C_4H_{11}N_2]^+,$ 100%); ESI⁻ $([C_{19}^{13}CH_{23}FN_4O_2-H]^-, 100\%), 255.9 ([C_{19}^{13}CH_{23}FN_4O_2-C_5H_{11}N_2O]^-, 100\%)$ 43%).

2-(2-(Ethylamino)ethyl)-1*H*-isoindole-1,3(2*H*)-dione hydrochloride (7)

A mixture of phthalimide (50.0 mmol, 7.36 g) and *N*-ethylethylenediamine (50.0 mmol, 4.41 g) was heated to 100°C for 3 h, followed by an additional 1 h at 130°C. Ethanol (100 mL) was added to the hot mixture and, upon cooling to room temperature, gaseous hydrogen chloride was introduced to obtain a white precipitate. The suspension was extended with ethanol (150 mL) and heated for recrystallization to obtain **7** as white needles (8.21 g, 64%), mp 236°C, lit.²⁰ 232–234°C. ¹H NMR (DMSO-*d*₆) δ : 1.18 (t, 3H, ³J (H,H) = 7.3 Hz, CH₂CH₃), 2.95 (app d, 2H, ³J (H,H) = 6.3 Hz, CH₂CH₃), 3.16 (app s, 2H, (CO)₂NCH₂CH₂), 3.91 (t, 2H, ³J (H,H) = 6.0 Hz, (CO)₂NCH₂CH₂), 7.78–7.90 (m, 4H, 4-H, 5-H, 6-H, 7-H), 9.21 (s, 2H, N⁺H₂); ¹³C NMR (DMSO-*d*₆) δ : 10.92 (CH₂CH₃), 34.03 ((CO)₂NCH₂CH₂), 41.83 (CH₂CH₃), 44.27 ((CO)₂NCH₂CH₂), 123.17 (C-4, C-7), 132.12 (C-3a, C-7a), 134.46 (C-5, C-6), 168.05 (C-1, C-3). HPLC purity: 95.8%.

N- (2 - ([²H₅]Diethylamino)ethyl)- 5-((5-fluoro-2-oxo-1,2-dihydro - 3H - indol-3-ylidene)methyl)-2,4-dimethyl-1H-pyrrole-3-carboxamide (8, ²H₅-SU11248)

A suspension of 2-(2-(ethylamino)ethyl)-1*H*-isoindole-1,3(2*H*)dione hydrochloride (**7**; 10.0 mmol, 2.55 g), $[^{2}H_{5}]$ ethyl iodide (12.0 mmol, 1.93 g) and potassium carbonate (20.0 mmol, 2.75 g) in acetonitrile (25 mL) was stirred 16 h at room temperature. The solvent was removed and the residue subjected to column chromatography on silica using ethyl acetate containing 5% triethylamine. Fractions containing 2-(2-([²H₅]diethylamino)ethyl)-1H-isoindole-1,3(2H)-dione were collected to obtain a viscious oil (5.5 mmol, 55%). Hydrazine (5.5 mmol, 0.28 g hydrazine hydrate, 100%) and ethanol (20 mL) were added and the reaction mixture refluxed for 1 h. The solvent was removed and the residue dissolved in N,N-dimethylformamide (10 mL). In separate flask, 5-((5-fluoro-2-oxo-1,2-dihydro-3H-indol-3а ylidene) methyl) - 2,4 - dimethyl - 1*H* - pyrrole - 3 - carboxylic acid (4a; 5.0 mmol, 1.51 g), 1-hydroxybenzotriazole (7.5 mmol, 1.00 g), 1-(3-dimethylaminopropyl)-3-ethylcarbodiimide were stirred in N,N-dimethylformamide (50 mL) for 2 h at room temperature. The solution prepared above was added and the mixture stirred for additional 16 h. Upon solvent removal, the residue was taken up in a mixture of ethyl acetate (100 mL) and methanol (10 mL), heated to reflux and filtered hot. The remaining solid was subjected to preparative HPLC on C18-silica using a gradient of methanol-water to afford 8 as a yellow powder (1.28 g, 63%), mp 217°C. ¹H NMR (DMSO- d_6) δ : 0.97 (t, 3H, ³J (H,H) = 7.1 Hz, CH₂CH₃), 2.41 (s, 3H, 2-CH₃), 2.43 (s, 3H, 4-CH₃), 2.52 (app q, 2H, ${}^{3}J$ (H,H) = 7.6 Hz, CH₂CH₃), 2.55 (app t, 2H, ${}^{3}J$ (H,H) = 6.7 Hz, CONHCH₂CH₂), 3.28 (app q, 2H, $(H,H) = 6.7 \text{ Hz}, \text{ CONHC}\underline{H}_2\text{CH}_2), 6.83 \text{ (dd, 1H, }^3J \text{ (H,H)} = 8.5 \text{ Hz}, {}^4J$ $(H,F) = 4.4 Hz, 7'-H), \overline{6.91} (ddd, 1H, {}^{3}J (H,F) = 9.0 Hz,$ (H,H) = 9.0 Hz, ⁴J (H,H) = 2.6 Hz, 6'-H), 7.40 (t, 1H, 31 (H,H) = 5.1 Hz, CONHCH₂CH₂), 7.70 (s, 1H, 3'-CH-5), 7.73 (dd, 1H, ${}^{3}J$ (H,F) = 9.5 Hz, ${}^{4}J$ (H,H) = 2.5 Hz, 4'-H), 10.85 (s, 1H, NH_{ind}), 13.66 (s, 1H, NH_{pvr}); ¹³C NMR (DMSO-*d*₆) δ: 10.71 (4-CH₃), 11.96 (CH₂<u>C</u>H₃, C²H₂<u>C</u>²H₃), 13.46 (2-CH₃), 37.07 (CONH<u>C</u>H₂CH₂), 46.63 (CH₂CH₃, C²H₂C²H₃), 51.75 (CONHCH₂CH₂), 106.03 (d, 1C, ²J (C,F) = 25.5 Hz, C-4'), 110.13 (d, 1C, ³J (C,F) = 8.4 Hz, C-7'), 112.48 (d, 1C, ²*J* (C,F) = 24.0 Hz, C-6'), 114.73 (d, 1C, ⁴*J* (C,F) = 3.0 Hz, C-3'), 120.85 (C-3), 124.98 (3'-CH-5), 125.92 (C-5), 127.30 (d, 1C, ³J (C,F)' = 9.4 Hz, C-3a'), 130.28 (C-4), 134.66 (C-7a'), 136.69 (C-2), 158.36 (d, 1C, ^{1}J (C,F) = 233 Hz, C-5'), 164.65 (CONHCH₂CH₂), 169.70 (C-2'). LC-MS: purity 99.8%. MS²: ESI⁺ 404.1 ($[C_{22}H_{22}^2H_5FN_4O_2+H]^+$, 36%), 326.1 ($[C_{22}H_{22}^2H_5FN_4O_2 C_4H_5^2H_5N]^+$, 49%), 282.9 ($[C_{22}H_{22}^2H_5FN_4O_2-C_6H_{10}^2H_5N_2]^+$, 100%); ESI^{-} 401.9 ($[C_{22}H_{22}^{2}H_{5}FN_{4}O_{2}-H]^{-}$, 100%), 254.9 $([C_{22} H_{22}^2 H_5 F N_4 O_2 - C_7 H_{10}^2 H_5 N_2 O_3]^-, 36\%).$

Conclusion

A concise five-step synthesis was developed to obtain either ¹³C- or ²H-labelled sunitinib (SU11248) or its primary metabolite (SU12662). All compounds were characterized in detail by NMR and/or LC-MS techniques; ¹³C NMR assignments of the sunitinib scaffold are reported for the first time. The reference compounds and internal standards were used to establish a LC-MS procedure that allows for the quantification of both compounds in human blood samples as a basis for PK/PD modelling studies of sunitinib, such as CESAR P-I-007. Details of which will be disclosed in a future communication.

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Supporting Information: HPLC chromatograms for compounds **1**, **2a-b**, **3a-b**, **4a-b**, and **7**, as well as LC-MS traces of compounds **5**, **6a-b**, and **8** are available at http://www.interscience.wiley.com.

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